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# ATMOSPHERIC COLD PLASMA AS NEW STRATEGY FOR FOODS PROCESSING - AN OVERVIEW

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**Abstract:** The atmospheric cold plasma it is a non-thermal processing method. This technique becomes a subject of high interest for a wide variety of technologies including the food industry. The atmospheric cold plasma it is proposed for decontamination of fruits and vegetables, especially, without changes in sensory attributes. This paper summarizes some relevant aspects of atmospheric cold plasma including generation, microbial inactivation mechanism and plasma applications in food industry. Although some aspects of antimicrobial mechanisms were presented here, more research should be done to clarify the antimicrobial mechanisms, in addition to confirm that no harmful by-products are generated by this technology.

Keywords: food, microbial inactivation, non-thermal processing.

#### Introduction

The foods may be contaminated by different microorganisms or can undergo deterioration from natural food enzymes. The pathogenic and spoilage microorganisms are problematic in the food industry due to their significant public health risks and economic impact (Stoica et al., 2011; Afshari and Hosseini, 2014). Therefore, in the food industry, the control of undesirable microorganisms is essential and decisive (Stoica et al., 2011). There are a lot of methods to destroy these microorganisms, such as: thermal technologies, e.g. sterilization, pasteurization, ohmic heating, etc. (Afshari and Hosseini, 2014) and non-thermal technologies, e.g. high hydrostatic pressure, pulsed electric fields, high voltage arc discharge (Stoica et al., 2011; Stoica et al., 2013; Afshari and Hosseini, 2014). On the other hand, the thermal technologies lead to unwanted changes in the foods' sensory attributes (by overheating) or to low nutritional value of the food products (Fernández et al., 2012; Stoica et al., 2013; Afshari and Hosseini, 2014), while the non-thermal technologies are often technically difficult to apply into production, expensive and require specialized equipment and trained personnel (Garcia-Gonzalez et al., 2007; Afshari and Hosseini, 2014; Misra et al., 2014b). ACP: atmospheric cold plasma, a nonthermal plasma technology, is proposed as a potential alternative to traditional methods for decontamination of foods (Bárdos and Baránková, 2010; Naïtali et al., 2010; Niemira, 2012; Afshari and Hosseini, 2014; Ziuzina et al., 2014). This technology does not require extreme process conditions and offers great opportunities for food product preservation (Fernández et al., 2012; Rod et al., 2012; Stoica et al., 2013), where the heat is not desirable for it (Afshari and Hosseini, 2014) in conjunction with maintenance of sensory attributes of the treated foods (Ziuzina *et al.*, 2014). This article briefly summarizes relevant aspects of ACP including plasma generation, microbial inactivation mechanism and plasma applications in food processing.

### Generation of plasma

The plasma is considered to be the forth state of matter (Tendero et al., 2006; Shakila Banu et al,. 2012; Suhem et al., 2013; Surowsky et al., 2013; Afshari and Hosseini, 2014; Mai-Prochnow et al., 2014). The plasma (or more correctly, the gas discharge plasma) is a mixture of partially ionized gas (Bárdos and Baránková, 2010, Niemira, 2012; Ryu et al., 2013; Pankaj et al,. 2014) that contains reactive species, such as: electrons, positive and negative ions, free radicals, gas atoms and photons (Bárdos and Baránková, 2010; Fernández et al., 2012; Stoica et al., 2013; Afshari and Hosseini, 2014; Pankaj et al., 2014). The reactive species generated by electron collision play a key role in the microbial plasma inactivation process (Mai-Prochnow et al., 2014). The ACP is dominated by reactive neutral species, such as: oxygen atoms, singlet oxygen and ozone rather than ions (Shibata et al., 1996; Schutze et al., 1998). In gas discharges at atmospheric pressure in air, the main reactive species are ozone, atomic oxygen, superoxide, peroxide, hydroxyl radicals (Afshari and Hosseini, 2014) with dominating role (Deng et al., 2006) and nitric oxide and nitrogen dioxide (Afshari and Hosseini, 2014). The reactive species can cause damages in proteins, lipids and nucleic acids (Cabiscol and Ros, 2000; Afshari and Hosseini, 2014; Mai-Prochnow et al., 2014). There are several means for artificial plasmas generation. The plasma is generated when the energy (Niemira, 2012; Shakila Banu et al., 2012) is applied across a dielectric gas or fluid (Shakila Banu et al., 2012). Any kind of energy can be used for gas ionization: microwave, radio frequency, electric or electromagnetic field, thermal, optical, radioactive and X-rays (Bárdos and Baránková, 2010; Afshari and Hosseini, 2014; Pankaj et al., 2014). The most useful ionization tools are electric or electromagnetic fields. When it is applied a voltage of about 30 kV/cm between electrodes (cathode and anode) separated by 1 cm in ambient air, electric breakdown and ignition of atmospheric air plasma can occur. Such electric breakdown can have the form of discrete sparks, but under certain conditions it is also possible to create a uniform-looking steady glow-type air discharge. The properties of plasma generally depend on the power, type of power (alternating current, direct current, pulsed. frequency) and type of gas (Bárdos and Baránková, 2010). The plasmas can be generated at low and high gas pressures (Bárdos and Baránková, 2010; Afshari and Hosseini, 2014). In plasmas generated at low gas pressures, where the density of gas particles available for ionization is low and the collision frequency is low, electron energies remain high compared to ion energies (is the case of the cold plasma, non-equilibrium plasma) (Bárdos and Baránková, 2010). The ACP, which is the most common plasma (Moisan et al., 2001; Bárdos and Baránková, 2010), it is said "cold" because the temperature in plasma reactor stays the near room temperature (Shakila Banu et al., 2012). Conversely, the high gas pressures with correspondingly high collision frequencies lead to equilibrium in the plasma (is the case of the hot plasma). The most feasible way to create non-equilibrium plasmas is "pumping" of power selectively to electrons (Bárdos and Baránková, 2010). The gas breakdown in a stationary electric field between electrodes it is the same of the production of electrons in a simple diode system. An electron emitted from the cathode collides along a unit length with a neutral gas particle and forms an ion and an additional (initial) electron (Bárdos and Baránková, 2010). The positive ions are attracted to the cathode and upon impact form secondary electrons which take part in subsequent ionizations in an avalanche type process. The secondary electron emission depends on a number of factors, such as: the material and the temperature of the cathode, the pretreatment of the cathode surface, gas and the effects of energetic ultraviolet radiation. In a stationary electric field, due to high collision frequencies at high gas pressure, it is nearly impossible to generate nonequilibrium cold plasma. The electric breakdown usually starts as a transient discharge in the form of a spark and at higher powers it easily turns into a high-current hot arc. The gases can also be ionized by electromagnetic (oscillating) field having amplitude of the electric component high enough for breakdown. Due to the immensely different mobility's of ions and electrons at high frequencies, the alternating current fields enable pumping of the power selectively to electrons, thus enabling nonequilibrium cold plasma at high gas pressures (Bárdos and Baránková, 2010). The alternating current power can be radiated into the plasma with electrodeless discharges (without electrodes), or with electrodes according to similar principles for direct current breakdown. The breakdown voltage depends on the gas, decreases with alternating current frequency due to a change in the electron loss mechanism. At high gas pressures, the electrons make many collisions per oscillation, and the production of secondary electrons at electrode surfaces is also less important. The magnitude of the electric field and the phase of the electron motion have a governing effect (Bárdos and Baránková, 2010). Often the noble gases are employed for inducing plasmas, but this can increase the cost of treatments. Thus, for inducing plasma should be the use of ambient air (Misra et al., 2014a).

#### Microbial inactivation mechanism of plasma

The plasma treatment can effectively inactivate a wide range of microorganisms (De Geyter and Morent, 2012; Fernández et al., 2012; Ryu et al., 2013; Wolf, 2013; Afshari and Hosseini, 2014) (prokaryotic and eukaryotic), (Li et al., 2013; Ryu et al., 2013; Mai-Prochnow et al., 2014) including spores and viruses (Li et al., 2013; Afshari and Hosseini, 2014; Mai-Prochnow et al., 2014). In many cases, the prokaryotic microorganisms (bacteria) are more vulnerable to plasma (Park et al., 2004; Lee et al., 2006; Muranyi et al., 2007; Tang et al., 2008; Kamgang-Youbi et al., 2009; Xiong et al., 2010). Bacteria from biotic and abiotic surfaces are effective killed by plasma (Baik et al., 2013; Pavlovich et al., 2013) at different rates and that the different death rates are probably caused by differential changes in components and ions in the solutions (Baik et al., 2013). The plasma generates different level of lethal effects on microorganisms through interaction with microbial surrounding environments (water, pH, nutrients, osmotic stability and temperature) (Ryu et al., 2013). The use of plasma as a sterilization method was first patented in 1968 and the plasma made from oxygen was first applied in 1989 (Afshari and Hosseini, 2014). After

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1990s, considerable researches have been done on mechanisms of microbial inactivation by plasma (Noriega et al., 2011; Shakila Banu et al., 2012; Ryu et al., 2013; Afshari and Hosseini, 2014), however, these researches are remained very limited (Ryu et al., 2013) and the exact mechanisms of this process remain unclear (Mai-Prochnow et al., 2014). The bacterial killing occurs via three different mechanisms: (a) direct permeabilization of the cell membrane or wall, leading to leakage of cellular components, including potassium, nucleic acids and proteins; (b) critical damage of intracellular proteins from oxidative or nitrosative species and (c) direct chemical nucleic acids damage (Mai-Prochnow et al., 2014). The reactive species can cause serious damages of the cells (Cabiscol and Ros, 2000; Afshari and Hosseini, 2014; Mai-Prochnow et al., 2014). Hydrogen peroxide was found to be the causative antibacterial agent that damages ironsulphurand mononuclear iron enzymes (Mai-Prochnow et al., 2014). The hydroxyl radicals (OH<sup>-</sup>) in plasma have toxic effects (Kang et al., 2011), being involved in membrane lipid peroxidation with the increase of membrane permeability (Ryu et al., 2013; Mai-Prochnow et al., 2014), through a destructive oxidation reactions (Gaunt et al., 2006) (equations 1 - 4) leading to unsaturated lipids disintegrating into lipid peroxides (Surowsky et al., 2013).

$$L + OH \cdot \rightarrow L \cdot + H_2O \qquad (eq.1)$$

$$L \cdot + O_2 \rightarrow L - OO \cdot$$
 (eq. 2)

$$L - 00 \cdot + L \rightarrow L \cdot + L - 00H (eq. 3)$$

$$L - OOH \to L - O \cdot$$
 (eq. 4)

The ozone can accelerate the chain reaction by oxidation and leads to a reduction of membrane fluidity (Gaunt *et al.*, 2006) and the functions of the membrane lipids are compromised. The lipid peroxidation generates the stables aldehydes which can travel long distance to attack targets (Gaunt *et al.*, 2006). The aldehydes are very reactive and can damage the proteins (Singh and Singh, 1982). In case of proteins, the reactive species react with the amino-acid chains and cause proteins structure

changes (Stadtman, 1992; Berlett and Stadtman, 1997; Cabiscol and Ros, 2000; Surowsky et al., 2013) and damage the cells (Afshari and Hosseini, 2014) and the spores (Laroussi, 2005; Afshari and Hosseini, 2014). Nucleic acids can be damaged, also (Farr and Kogoma, 1991; Afshari and Hosseini, 2014). In ACP, the reactive species are thought to be the major components that have antimicrobial effects (Shintani et al., 2010; Afshari and Hosseini, 2014), while the effect of ultraviolet irradiation is The minor (Afshari and Hosseini, 2014). inactivation microorganism mechanisms depend on: type of food product, ACP design, type of microorganisms, voltage, pressure, gas gas composition, water content in the gas, and distance of the microorganism from the discharge glow (Afshari and Hosseini, 2014; Ziuzina et al., 2014). The microbial response also depends on the bacterial mode of growth, with bacteria growing in biofilms requiring a longer exposure time before becoming inactivated (Mai-Prochnow et al., 2014).

### Atmospheric cold plasma applications

Due to their design simplicity and low operating cost, this form of discharge plays an important role for many technological applications. Several ACP applications have been identified in literature (Law et al., 2012), such as: food industry (Fernández et al., 2012; Shakila Banu et al., 2012; Surowsky et al., 2013; Pankaj et al., 2014; Ziuzina et al., 2014), medical and clinical (Terrier et al., 2009; Isbary et al., 2013; Hoffmann et al., 2013; Arora et al., 2014; Mai-Prochnow et al., 2014), materials processing (Law et al., 2012; Callard Preedy et al., 2014; Sato et al., 2014), material analysis (Sato et al., 2014), surface modification (Law et al., 2012; Sato et al., 2014; Pankaj et al., 2014), light source (Law et al., 2012; Sato et al., 2014) and microplasma chip (Sato et al., 2014). ACP technology offers distinct advantages for decontamination of foods (Misra et al., 2014b). ACP is increasingly under research for decontamination of fresh produce (Kabir Jahid et al., 2014; Ziuzina et al., 2014) fruits and vegetables, especially (Klockow et al., 2009; Fan et al., 2012; Fernández et al., 2012; Baier et al., 2014; Misra et al., 2014a; Ziuzina et al., 2014). ACP technology can be used to treat a variety of vegetables: fresh tomatoes (Bermúdez-Aguirre et al., 2013; Misra et al. 2014a; Pankaj et al., 2014), cherry tomatoes (Misra et al., 2014a), lettuce (Shakila Banu et al., 2012; Bermúdez-Aguirre et al., 2013; Kabir Jahid et al., 2014; Misra et al., 2014a), carrots (Bermúdez-Aguirre et al., 2013; Misra et al., 2014a), cucumbers and broccoli. The tomatoes and the lettuce were easier to decontaminate than the carrots, probably because of the surface structure (Bermúdez-Aguirre et al., 2013). Also, ACP is used to reduce the microorganisms on strawberries (Misra et al., 2014a), apples (Shakila Banu et al., 2012), melons and mangos (Shakila Banu et al., 2012), pears, spice e.g. red pepper (Kim et al., 2014), nuts (Shakila Banu et al., 2012). ACP can successfully decontaminate of the fresh meat and poultry (Shakila Banu et al., 2012) and meat products (Noriega et al., 2011; Frohling et al., 2012; Rod et al., 2012), e.g. bacon (Kim et al., 2011), ham (Shakila Banu et al., 2012) and ready to eat meat (Rod et al., 2012) and cheese (Shakila Banu et al., 2012). ACP it is able to control the microorganisms in the cereal industry (Shakila Banu et al., 2012; Suhem et al., 2013) because it doesn't generate too much heat which means it may damage food nutrition less (Suhem et al., 2013). Recently, some researches show that ACP is suitable for inactivate enzymes from fruit or vegetable sources (Pankaj et al., 2014). ACP is a powerful tool for surface decontamination of not only foods but also food packaging materials (Pankaj et al., 2014; Surowsky et al., 2014) (plastic bottles, lids and films) without adversely affecting their bulk properties, and does not result in any liquid effluents (residues) (Pankaj et al., 2014). Newer, ACP technology can be combined with the essential oil (e.g. clove oil) to decontaminate the cellulose-based food packaging (Matan et al., 2014).

## Conclusions

The atmospheric cold plasma, by far, is one of the newest technologies used in food industry for microbial inactivation. In the microbial inactivation plasma process, a key role is attributed to reactive species (atomic oxygen, singlet oxygen, ozone, superoxide, hydrogen peroxide, hydroxyl radicals, nitric oxide and nitrogen dioxide) generated by electron collision. The atmospheric cold plasma treatment can effectively inactivate a wide range of microorganisms including spores and viruses,

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through permeabilization of the cell membrane, damage of the intracellular proteins and the nucleic acids and the inactivate of some enzymes. This technology is considered to be very promising alternative to thermal new processing technologies and it has many interesting applications in the food industry: decontamination especially of fruits and vegetables, spice, nuts, raw and meat products, cheese, or decontamination of the food packaging materials and newer can be combined with some antimicrobial agents such as essential oils. However, more information's are needed to clarify the microbial inactivation mechanisms and to confirm that no harmful by-products are generated by atmospheric cold plasma treatment.

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## References

- Afshari R., Hosseini H. (2014) Non-thermal plasma as a new food preservation method, Its present and future prospect, *Journal of Paramedical Sciences*, 5 (1), 2008-4978.
- Arora V., Nikhil V., Suri N.K., Arora P. (2014) Cold Atmospheric Plasma (CAP) in Dentistry, *Dentistry*, 4 (1), 189-193.
- Baier M., Görgen M., Ehlbeck J., Knorr D., Herppich W.B., Schlüter O. (2014) Non-thermal atmospheric pressure plasma: Screening for gentle process conditions and antibacterial efficiency on perishable fresh produce, *Innovative Food Science and Emerging Technologies*, 22, 147–157.
- Baik K.Y., Kim Y.H., Ryu Y.H., Kwon H.S., Park G., Uhm H.S., Choi E.H. (2013) Feeding-Gas Effects of Plasma Jets on *Escherichia coli* in Physiological Solutions, *Plasma Processes and Polymers*, 10 (3), 235–242.

- Bárdos L., Baránková H. (2010) Cold atmospheric plasma: Sources, processes, and applications, *Thin Solid Films*, 518, 6705–6713.
- Berlett B.S. Stadtman E.R. (1997) Protein oxidation in aging, disease, and oxidative stress, *Journal of Biological Chemistry*, 272 (33), 20313-20316.
- Bermúdez-Aguirre D., Wemlinger E., Pedrow P., Barbosa-Cánovas G., Garcia-Perez M. (2013) Effect of atmospheric pressure cold plasma (APCP) on the inactivation of *Escherichia coli* in fresh produce, *Food Control*, 34 (1), 149-157.
- Cabiscol E., Tamarit J., Ros, J. (2000) Oxidative stress in bacteria and protein damage by reactive oxygen species, *International Microbioly*, 3 (1) 3-8.
- De Geyter N., Morent R. (2012) Nonthermal Plasma Sterilization of Living and Nonliving Surfaces, *Annual Review of Biomedical Engineering*, 14, 255–274.
- Deng X.T., Shi J.J., Kong M.G. (2006) Physical mechanisms of inactivation of *Bacillus subtilis* spores using cold atmospheric plasmas, *IEEE Transactions* on *Plasma Science*, 34 (4), 1310-1316.
- Fan X., Sokorai K.J., Engemann J., Gurtler J.B., Liu Y. (2012) Inactivation of *Listeria innocua*, *Salmonella Typhimurium*, and *Escherichia coli* O157:H7 on surface and stem scar areas of tomatoes using inpackage ozonation, *Journal of Food Protection*, 75 (9), 1611-1618.
- Farr S.B., Kogoma T. (1991) Oxidative stress responses in *Escherichia coli* and *Salmonella typhimurium*, *Microbiological Reviews*, 55 (4), 561-585.
- Fernández A., Shearer N., Wilson D.R., Thompson A. (2012.) Effect of microbial loading on the efficiency of cold atmospheric gas plasma inactivation of *Salmonella enterica serovar Typhimurium*, *International Journal of Food Microbiology*, 152 (3), 175–180.
- Frohling A., Durek J., Schnabel U., Ehlbeck J., Bolling J., Schlüter O. (2012) Indirect plasma treatment of fresh pork: decontamination efficiency and effects on quality attributes, *Innovative Food Science & Emerging Technologies*, 16, 381-390.
- Garcia-Gonzalez L., Geeraerd A.H., Spilimbergo S., Elst K., Van Ginneken L., Debevere J., Van Impe J.F., Devlieghere F. (2007) High pressure carbon dioxide inactivation of microorganisms in foods: The past, the

present and the future, *International Journal of Food Microbiology*, 117 (1), 1–28.

- Gaunt L.F., Beggs C.B., Georghiou G.E. (2006) Bactericidal action of the reactive species produced by gas-discharge nonthermal plasma at atmospheric pressure: A review, *IEEE Transactions on Plasma Science*, 34 (4), 1257-1269.
- Hoffmann C., Berganza C., Zhang J. (2013) Cold Atmospheric Plasma: methods of production and application in dentistry and oncology, *Medical Gas Research*, 3 (1), 21.
- Isbary G., Stolz W., Shimizu T., Monetti R., Bunk W., Schmidt H.-U., Morfill G.E., Klämpfl T.G., Steffes B., Thomas H.M., Heinlin J., Karrer S., Landthaler M., Zimmermann J.L. (2013) Cold atmospheric argon plasma treatment may accelerate wound healing in chronic wounds: Results of an open retrospective randomized controlled study in vivo, *Clinical Plasma Medicine*, 1 (2), 25–30.
- Kabir Jahid I., Han N., Ha S.D. (2014) Inactivation kinetics of cold oxygen plasma depend on incubation conditions of *Aeromonas hydrophila* biofilm on lettuce, *Food Research International*, 55, 181–189.
- Kamgang-Youbi G., Herry M., Meylheuc T., Brisset J.L., Bellon-Fontaine M.N., Doubla A., Naïtali M. (2009) Microbial inactivation using plasma-activated water obtained by gliding electric discharges, *Letters in Applied Microbiology*, 48 (1), 13–18.
- Kang S.K., Choi M.Y., Koo I.G., Kim P.Y., Kim Y., Kim G.I., Mohamed A-A.H., Collins G.I., J.K. (2011) Reactive hydroxyl radical-driven oral bacterial inactivation by radio frequency atmospheric plasma, *Applied Physics Letters*, 98, 143702 - 143702-3.
- Kim B., Yun H., Jung S., Jung Y., Jung H., Choe W., Jo C. (2011) Effect of atmospheric pressure plasma on inactivation of pathogens inoculated onto bacon using two different gas compositions, *Food Microbiology*, 28 (1), 9-13.
- Kim J.E., Lee D-U., Min S.K. (2014) Microbial decontamination of red pepper powder by cold plasma, *Food Microbiology*, 38, 128-136.
- Klockow P.A., Keener K.M. (2009) Safety and quality assessment of packaged spinach treated with a novel ozone-generation system, *LWT - Food Science and Technology*, 42 (6), 1047-1053.

- Laroussi M. (2005) Low temperature plasma-based sterilization: Overview and state of-the-art, *Plasma Processes and Polymers*, 2 (5), 391-400.
- Law V.J., O'Neill F.T., Twomey B., Milosavljevi V., Kong M.G., Anghel S.D., Dowling, D.P. (2012) Electrical power dissipation within a helium APPJ flowing afterglow and its impact on spatial-temporal properties, *IEEE Transitions on plasma science*, 40 (11), 2994-3002.
- Lee K., Paek K.H., Ju W.T., Lee Y. (2006) Sterilization of bacteria, yeast, and bacterial endospores by atmospheric-pressure cold plasma using helium and oxygen, *Journal Microbiology*, 44 (3), 269–275.
- Li Y.F., Taylor D., Zimmermann J.L., Bunk W., Monetti R., Isbary G., Boxhammer V., Schmidt H.U., Shimizu T., Thomas H.M., Morfill G.E. (2013) In vivo skin treatment using two portable plasma devices: Comparison of a direct and an indirect cold atmospheric plasma treatment, *Clinical Plasma Medicine*, 1 (2), 35–39.
- Mai-Prochnow A., Murphy A.B., McLean M., Kong M.G., Ostrikov K. (2014) Atmospheric pressure plasmas: Infection control and bacterial responses, *International Journal of Antimicrobial Agents*, 43 (6), 508–517.
- Matan N., Nisoa M., Matan N., Aewsiri T. (2014) Effect of cold atmospheric plasma on antifungal activities of clove oil and eugenol against molds on areca palm (Areca catechu) leaf sheath, *International Biodeterioration & Biodegradation*, 86 (Part C), 196-201.
- Misra N.N., Keener K.M., Bourke P., Mosnier J.P., Cullen P.J. (2014a). In-package atmospheric pressure cold plasma treatment of cherry tomatoes, *Journal of Bioscience and Bioengineering*, xx (1-6), http://dx.doi.org/10.1016/j.jbiosc.2014.02.005.
- Misra N.N., Patil S., Moiseev T., Bourke P., Mosnier J.P., Keener K.M., Cullen P.J. (2014b) In-package atmospheric pressure cold plasma treatment of strawberries, *Journal of Food Engineering*, 125, 131– 138.
- Moisan M., Barbeau J., Moreau S., Pelletier J., Tabrizian M., Yahia L.H. (2001) Low-temperature sterilization using gas plasmas: a review of the experiments and an analysis of the inactivation mechanisms, *International Journal of Pharmaceutics*, 226 (1-2), 1-21.

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- Muranyi P., Wunderlich J., Heise M. (2007) Sterilization efficiency of a cascaded dielectric barrier discharge, *Journal Applied Microbiology*, 103 (5), 1535–1544.
- Naïtali M., Kamgang-Youbi G., Herry J.M., Bellon-Fontaine M.N., Brisset J.L. (2010) Combined Effects of Long-Living Chemical Species during Microbial Inactivation Using Atmospheric Plasma-Treated Water, Applied Environmental Microbiology, 76 (22), 7662–7664.
- Niemira B.A. (2012) Cold Plasma Decontamination of Foods, *Annual Review of Food Science and Technology*, 3, 125-142.
- Noriega E., Shama G., Laca A., Díaz M., Kong M.G. (2011) Cold atmospheric gas plasma disinfection of chicken meat and chicken skin contaminated with *Listeria innocua*, *Food Microbiology*, 28 (7), 1293– 1300.
- Pankaj S.K., Bueno-Ferrer C., Misra N.N., Milosavljevi V., O'Donnell C.P., Bourke P., Keener K.M., Cullen P.J. (2014) Applications of cold plasma technology in food packaging, *Trends in Food Science & Technology*, 35 (1), 5-17.
- Park J.C., Park B.J, Han D.W., Lee D.H., Lee I.S., Hyun S.O., Chun M.S., Chung K.H., Aihara M., Takatori K. (2004) Fungal sterilization using microwave-induced argon plasma at atmospheric pressure, *Journal Microbiology Biotechnology*, 14, 188–192.
- Pavlovich M.J., Chen Z., Sakiyama Y., Clark D.S., Graves D.B. (2013) Effect of Discharge Parameters and Surface Characteristics on Ambient-Gas Plasma Disinfection, *Plasma Processes and Polymers*, 10 (1), 69–76.
- Preedy E.C., Brousseau E.B., Evans S.L., Perni S., Prokopovich P. (2014) Adhesive forces and surface properties of cold gas plasma treated UHMWPE, *Colloids and Surfaces A: Physicochemical and Engineering* Aspects, DOI: 10.1016/j.colsurfa.2014.03.052.
- Rod S.K., Hansen F., Leipold F., Knochel S. (2012) Cold atmospheric pressure plasma treatment of ready-to-eat meat: Inactivation of *Listeria innocua* and changes in product quality, *Food Microbiology*, 30 (1), 233-238.
- Ryu Y-H., Kim Y-H., Lee J-Y., Shim G-B., Uhm H-S., Park G., Choi E.H. (2013) Effects of Background Fluid on the Efficiency of Inactivating Yeast with

Non-Thermal Atmospheric Pressure Plasma, *PLoS ONE*, 8(6): e66231. doi: 10.1371/journal. pone. 0066231

- Sato R., Yasumatsu D., Kumagai S., Takeda K., Hori M., Sasaki M. (2014) An atmospheric pressure inductively coupled microplasma source of vacuum ultraviolet light, *Sensors and Actuators A: Physical*, 215 (15), 144–149.
- Schutze A., Jeong J.Y., Babayan S.E., Park J., Selwyn G.S., Hicks R.F. (1998) The atmospheric-pressure plasma jet: A review and comparison to other plasma sources, *IEEE Transactions on Plasma Science*, 26 (6), 1685-1694.
- Shakila Banu M., Sasikala P., Dhanapal A., Kavitha V., Yazhini G., Rajamani L. (2012) Cold plasma as a novel food processing technology, *International Journal of Emerging trends in Engineering and Development*, 4 (2), 803-818.
- Shibata M., Nakano N., Makabe T. (1996) Effect of O-2(a(1)Delta(g)) on plasma structures in oxygen radio frequency discharges, *Journal of Applied Physics*, 80 (11), 6142-6147.
- Shintani H., Sakudo A., Burke P., McDonnell G. (2010) Gas plasma sterilization of microorganisms and mechanisms of action, *Experimental and Therapeutic Medicine*, 1 (15), 731–738.
- Singh A., Singh, H. (1982) Time-scale and nature of radiation biological damage approaches to radiation protection and post-irradiation therapy, *Progress in Biophysics & Molecular Biology*, 39 (2), 69-107.
- Stadtman E.R. (1992) Protein oxidation and aging, *Science*, 257 (5074), 1220-1224.
- Stoica M., Bahrim G., Cârâc, G. (2011) Factors that Influence the Electric Field Effects on Fungal Cells. In: Méndez-Vilas A. (ed.): Science against microbial pathogens: communicating current research and technological advances, 291-302. Formatex Research Center, Badajoz.
- Stoica M., Mihalcea L., Borda D., Alexe P. (2013) Nonthermal novel food processing technologies. An overview, *Journal of Agroalimentary Processes and Technologies*, 19 (20), 212-217.
- Suhem K., Matan N., Nisoa M., Matan N. (2013) Inhibition of *Aspergillus flavus* on agar media and brown rice cereal bars using cold atmospheric plasma

treatment, International Journal of Food Microbiology, 161, 107–111.

- Surowsky B., Fischer A., Schlueter O., Knorr D. (2013) Cold plasma effects on enzyme activity in a model food system, *Innovative Food Science and Emerging Technologies*, 19, 146–152.
- Surowsky B., Fröhling A., Gottschalk N., Schlüter O., Knorr D. (2014) Impact of cold plasma on Citrobacter freundii in apple juice: Inactivation kinetics and mechanisms, International *Journal of Food Microbiology*, 174C, 63–71.
- Tang Y.Z., Lu X.P., Laroussi M., Dobbs F.C. (2008) Sublethal and killing effects of atmospheric-pressure, Nonthermal plasma on eukaryotic microalgae in aqueous media, *Plasma Processes and Polymers*, 5 (6), 552–558.
- Tendero C., Tixier C., Tristant P., Desmaison J., Leprince P. (2006) Atmospheric pressure plasmas: A review, *Spectrochimica Acta Part B: Atomic Spectroscopy*, 61(1), 2-30.

- Terrier O., Essere B., Yver M., Barthélémy M., Bouscambert-Duchamp M., Kurtz P., VanMechelen D., Morfin F, Billaud G., Ferraris O., Lina B., Rosa-Calatrava M., Moules V. (2009) Cold oxygen plasma technology efficiency against different airborne respiratory viruses, *Journal of Clinical Virology*, 45 (2), 119–124.
- Wolf R.A. (2013) Atmospheric Plasma Surface Antimicrobial Effects. In: Wolf RA (ed.): Atmospheric Pressure Plasma for Surface Modification, 181–194. John Wiley & Sons, Hoboken.
- Xiong Z., Lu X.P., Feng A., Pan Y., Ostrikov K. (2010)
  Highly effective fungal inactivation in He+
  O<sub>2</sub> atmospheric-pressure nonequilibrium plasmas, *Physics of Plasmas*, 17 (12), 123502-123502-6.
- Ziuzina D., Patil S., Cullen P.J., Keener K.M., Bourke P. (2014) Atmospheric cold plasma inactivation of Escherichia coli, Salmonella enterica serovar Typhimurium and Listeria monocytogenes inoculated on fresh produce, Food Microbiology, 42, 109-116.